

9-30-2022

Metabolite Signature of Fresh and Long-term Stored Coffee Pulp and Husk

Wahyu Lestari

Faculty of Medicine, Universitas Syiah Kuala, Aceh 23111, Indonesia, wahyu_lestari2000@unsyiah.ac.id

Kartini Hasballah

Department of Pharmacology, Faculty of Medicine, Universitas Syiah Kuala, Aceh 23111, Indonesia

Muhammad Yulianto Listiawan

Department of Dermatology and Venereology, Faculty of Medicine, Universitas Airlangga/Dr. Soetomo Hospital, Surabaya 60115, Indonesia

Sofia Sofia

Department of Biochemistry, Faculty of Medicine, Universitas Syiah Kuala, Aceh 23111, Indonesia

Follow this and additional works at: <https://scholarhub.ui.ac.id/science>

 Part of the [Food Chemistry Commons](#)

Recommended Citation

Lestari, Wahyu; Hasballah, Kartini; Listiawan, Muhammad Yulianto; and Sofia, Sofia (2022) "Metabolite Signature of Fresh and Long-term Stored Coffee Pulp and Husk," *Makara Journal of Science*: Vol. 26: Iss. 3, Article 8.

DOI: 10.7454/mss.v26i3.1354

Available at: <https://scholarhub.ui.ac.id/science/vol26/iss3/8>

This Article is brought to you for free and open access by the Universitas Indonesia at UI Scholars Hub. It has been accepted for inclusion in Makara Journal of Science by an authorized editor of UI Scholars Hub.

Metabolite Signature of Fresh and Long-term Stored Coffee Pulp and Husk

Wahyu Lestari^{1,2*}, Kartini Hasballah³, Muhammad Yulianto Listiawan⁴, and Sofia Sofia⁵

1. Faculty of Medicine, Universitas Syiah Kuala, Aceh 23111, Indonesia

2. Department of Dermatology and Venereology, General Hospital of Dr. Zainoel Abidin, Aceh 24415, Indonesia

3. Department of Pharmacology, Faculty of Medicine, Universitas Syiah Kuala, Aceh 23111, Indonesia

4. Department of Dermatology and Venereology, Faculty of Medicine, Universitas Airlangga/Dr. Soetomo Hospital, Surabaya 60115, Indonesia

5. Department of Biochemistry, Faculty of Medicine, Universitas Syiah Kuala, Aceh 23111, Indonesia

*E-mail: wahyu_lestari2000@unsyiah.ac.id

Received April 19, 2022 | Accepted September 22, 2022

Abstract

Every product from the food and agriculture industry produces waste that can cause environmental pollution when carelessly discarded. Coffee husk and pulp are the main wastes generated by coffee processing. The secondary metabolites in these wastes can still be utilized, but their level can be affected by long storage. This research aims to determine the differences in the compounds obtained from old and fresh Gayo Arabica coffee pulp and husk. Coffee husk and pulp samples from Gayo Arabica coffee are extracted for GC-MS analysis to determine similarities between fresh and long-term stored Gayo Arabica coffee husk and pulp. Results show differences in level and type among the secondary metabolites. Among various compounds, caffeine is found in all the samples. The compounds obtained from Gayo Arabica coffee husk and pulp, such as caffeine, (Z,Z)-9,12-octadecadienoic acid, and palmitic acid, can be used in health and agriculture.

Keywords: gayo arabica coffee, GC-MS, husk, pulp

Introduction

The term coffee is derived from “qahva” or “qahwa,” which means a drink made from plants. Initially, coffee cherry was consumed, and the seeds were crushed and mixed with animal fat to make energy-boosting supplements. Currently, 100 species of *Coffea* genus are found in tropical and subtropical regions. Two types of coffee, *Coffea arabica* and *Coffea canephora*, are commonly found in Indonesia [1, 2].

The largest producer of arabica coffee is in Northern Indonesia, namely, Gayo Lues, the central part of Aceh Province. The coffee produced in this area is called Gayo coffee. Cupping test ascribed a score of more than 80 for Gayo coffee, making it a specialty coffee [3]. The optimum condition for coffee growth can be found in the highland tropical region with altitude of 1.000–1.700 MASL and temperature of 18 °C–23 °C [4]. From a geographic perspective, Gayo highland fulfills this optimum condition [5]. Hence, the secondary metabolites from Gayo Arabica coffee are better than those from other types of coffee [6].

A natural product is a compound isolated from a living organism and is produced from primary and secondary

metabolites. A metabolite is an intermediate or product of metabolism and can be classified as primary or secondary. Primary metabolites directly affect growth, development, and reproduction. Secondary metabolites do not directly affect these processes but are still important for plants. The most bioactive metabolites are alkaloids, tannins, flavonoids, and phenolic compounds [7]. Secondary metabolites in plants can be an alternative source for drug discovery [8].

The anatomy of the coffee fruit consists of the outer skin, pulp (mesocarp), pectin layer, parchment coat, silver skin, bean, and center cut [5]. Coffee processing is divided into two parts: dry and wet processing [9, 10]. Almost half of the coffee fruit will become waste. The husk will be produced in the dry process, and the pulp is the main waste from the wet process [11]. The coffee husk contains polyphenols, anthocyanins, vitamin C, and some secondary metabolites, such as caffeine, alkaloids, and tannins. Coffee pulp contains 4%–12% proteins, 1%–2% fats, 6%–10% minerals, and 45%–89% carbohydrates [12]. Ameca *et al.* [13] identified eight phenolic acids in coffee pulp: hydroxybenzoic acid, chlorogenic acid, ferulic acid, caffeine acid, syringic acid, gallic acid, vanillic acid, and coumaric acid. The secondary metabolites of coffee can inhibit metabolic

diseases, such as diabetes, obesity, hypertension, and neural diseases [14–18].

The storage period can affect the secondary metabolites in plants. A previous study on broccoli observed a reduction in vitamin C levels with prolonged storage time because of the increase in vitamin C-degrading enzyme activity during storage [19]. Another study on oat plants reported a consistent result of decreasing secondary metabolite concentration (total phenolic content and caffeic acid) with prolonged storage time [20].

As a compound targeted to produce a beneficial effect on health, any changes on secondary metabolites are crucial. Reduced secondary metabolites will not have an optimum impact on health, despite being extracted from the same plants. This study aims to identify differences among the secondary metabolites of husk and pulp of Gayo Arabica coffee under fresh condition and long storage.

Methods

Plant Materials. Husks and pulps of Gayo Arabica coffee were collected from arabica coffee plants (*Coffea arabica* L.) in Gayo Lues, Central Aceh, Indonesia. The variety of the coffee is Ateng super, which has a stem height of 1.5 m and planting height of 1.450 MASL (Mean Above Sea Level). Fresh Gayo Arabica husks and pulps were harvested in December 2021. Long-term stored husks and pulps were obtained from coffee that has been stored for 1 year (December 2020). This research was conducted in the chemical engineering faculty of Universitas Syiah Kuala, Aceh, Indonesia.

Extraction and Gas Chromatography Mass-Spectrophotometry (GC-MS). Husks and pulps of Gayo Arabica coffee were extracted by maceration using absolute n-hexane. Liquid extract was evaporated using a rotary evaporator at 40 °C for further analysis. The secondary metabolites of Gayo Arabica coffee husk and pulp were analyzed using GC with an Rtx-5MS column (30 cm length). In brief, 1 g of evaporated crude extract was dissolved with 3 ml of ethanol absolute and vortex for 3 minutes with an extension of 15 minutes. The homogenates were filtered using 0.45 µl filter paper, and 1 µL of the filtrate was used for injection at GC-MS (Shidmazu, QP2010) under the following conditions: temperature of 280 °C, splitless injector mode, column temperature of 40 °C, retention time of 40 min, and detector temperature of 280 °C. Helium was used as the carrier gas, and the pressure, total flow rate, column flow, and split ratio were set at 4,3367 psi, 8.4 mL/min, 0.9 mL/min, and 5:1, respectively. The MS was set to scan mode with a scan range of 30–500 m/z, ion source temperature of 200 °C, interface temperature of 300 °C, solvent cut time of 2.5 minutes, event time of 0.3 seconds, and a scan speed of 1666. Compounds were

identified by comparing the obtained mass spectra of each compound with those from the WILLEY7 database. [2]

Results and Discussions

Several methods, such as GC-MS, Fourier transform infrared (FTIR) spectroscopy, and high-performance liquid chromatography (HPLC), are commonly employed for the detection of secondary metabolites. HPLC is not usual applied in detecting secondary metabolites, and FITR is not available in Aceh. Hence, GC-MS was selected for this study. Comparison of total ion chromatogram between fresh and long-term stored Gayo Arabica coffee husk is shown in Figure 1 and Figure 2. Differences were observed between the fresh and stored husk samples. The total compounds detected were 17, and 9 of which were not identified with different retention times shown in Table 1 and Table 2.

The first retention time of fresh husk was 11.194 minutes and the last was 34.168 minutes with total compounds 1.935% and 1.333% respectively. While the long-term stored husk showed the first retention time was 11.183 minutes and the last 24.321 minutes with total compound 3.265% and 4.394% respectively.

Differences of the compound were found between fresh and stored husk that shown in Table 3. Hydroxymethylfurfural (HMF) and (Z,Z)-9,12-octadecadienoic acid were the compounds detected only in long-term stored husks. The formation of HMF depends on temperature, pH, and food composition, so this compound is often used as a marker of food quality. HMF is formed during heating through the Maillard reaction and sugar degradation [21]. Owing to the sugar content of the coffee husk, storage causes sugar degradation and HMF formation [22]. GC-MS analysis revealed that the fatty acids can be classified as saturated or unsaturated. The compounds included in the saturated fatty acids are palmitic acid and stearic acid. Linoleic acid and (Z,Z)-9,12-octadecadienoic acid belong to the unsaturated fatty acids. Saturated fatty acids are susceptible to oxidation during storage, and unsaturated fatty acids are resistant to oxidation [23].

Figures 3 and 4 show the comparison of the total ion chromatogram of the fresh and long-term stored pulps of Gayo Arabica coffee. Some differences in retention time and number of detected compounds were noted between the two samples. Among the 35 compounds detected, 7 were not identified. Figure 5 show three major compounds identified with the biggest area, caffeine, hexadecenoic acid, and elaidic acid. Each compound has retention time 15.363 minutes, 16.566 minutes, and 19.05 minutes sequentially. Caffeine was detected in the fresh and long-term stored pulp samples. The retention time of caffeine was 15.363 minutes in the fresh pulp and 21.48

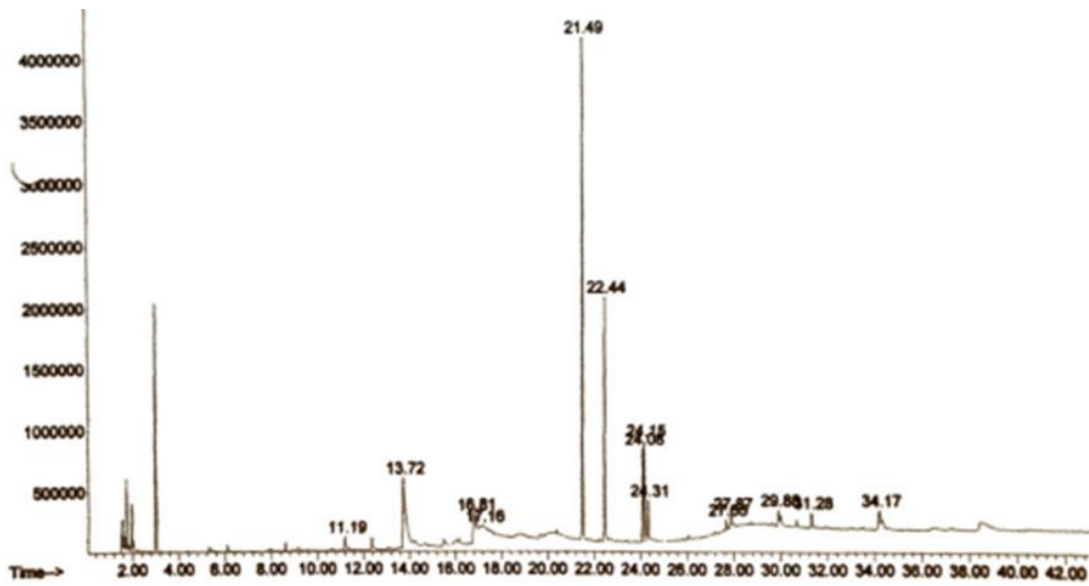


Figure 1. Total Ion Chromatogram of the Fresh Husk of Gayo Arabica Coffee

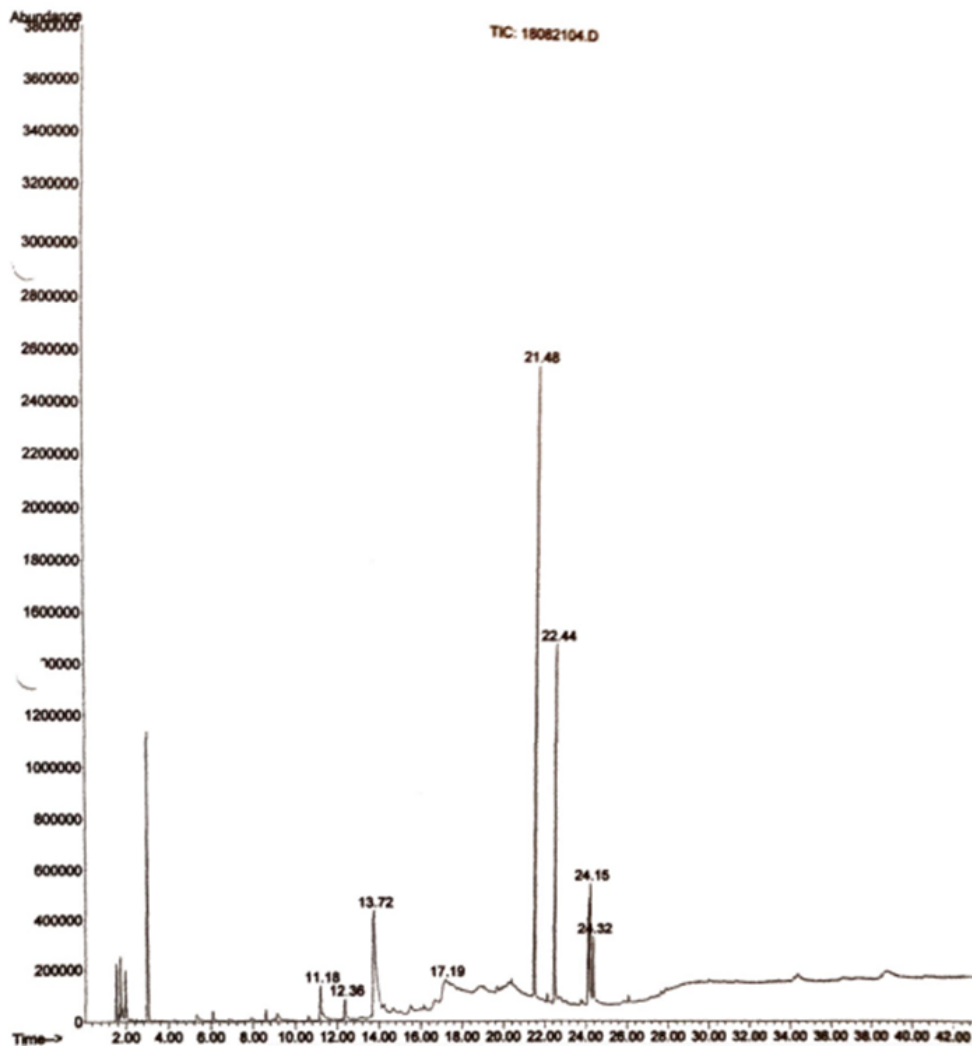


Figure 2. Total Ion Chromatogram of the Long-term Stored Husk of Gayo Arabica Coffee

Table 1. Mass Selective Detector (MSD) of the Fresh Husk of Gayo Arabica Coffee

Peak	Retention Time (Minute)	First Scan	Max Scan	Last Scan	PK TY	Peak Height	Corr. Area	Corr. % Max	% Of Total
1	11.194	1927	1941	1977	BB	113572	4998874	6.16%	1.935%
2	13.723	2317	2383	2460	BV	551336	44978770	55.42%	17.413%
3	16.813	2852	2923	2949	BV	204595	14193110	17.49	5.495%
4	17.168	2949	2985	3034	VB 9	72661	14986137	18.46%	5.802%
5	21.488	3705	3740	3783	BV	3985580	81161784	100.00%	31.421%
6	22.438	3890	3906	3942	BV	1933825	36135994	44.52%	14.990%
7	24.085	4175	4194	4199	BV	730854	14050215	17.31%	5.439%
8	24.148	4199	4205	4226	VV 2	803121	21456007	26.44%	8.306%
9	24.314	4226	4234	4267	VB	318991	8105089	9.99%	3.138%
10	27.645	4751	4816	4825	BV	83776	1632407	2.01%	0.632%
11	27.868	4825	4855	4896	VB 6	121133	6810521	8.39%	2.637%
12	29.882	5183	5207	5216	BV 3	96056	1684881	2.08%	0.652%
13	31.278	5421	5451	5483	BB	122636	4670221	5.75%	1.808%
14	34.168	5918	5956	5969	BV 3	114775	3443584	4.24%	1.333%

Table 2. MSD of Long-term Stored Husk of Gayo Arabica Coffee

Peak	Retention Time (Minute)	First Scan	Max Scan	Last Scan	PK TY	Peak Height	Corr. Area	Corr. % Max	% of Total
1	11.183	1923	1939	1983	BB	134371	5769343	11.67%	3.265%
2	12.362	2099	2145	2183	BB 3	78512	3644480	7.37%	2.063%
3	13.723	2334	2383	2461	BV	412014	38525802	77.90%	21.804%
4	17.185	2857	2988	3103	BB	74696	23317843	47.15%	13.197%
5	21.477	3687	3738	3797	BB	2540282	49456740	100.00%	27.991%
6	22.444	3872	3907	3942	BV	1406515	26769393	54.13%	15.151%
7	24.149	4172	4205	4227	BV 3	485776	21441406	43.35%	12.135%
8	24.321	4227	4235	4292	VB	246019	7763023	15.70%	4.394%

Table 3. Secondary Metabolites in the Fresh and Long-term Stored Husk of Gayo Arabica Coffee

Retention Time (Minutes)	Compound Name	Molecular Weight	% Area of Fresh Husk	% Area of Long-Term Stored Husk
11.18	Unknown	-	1.94	3.27
12.36	Unknown	-	Nd	2.06
13.72	HMF	126.11	Nd	21.80
13.73	Unknown	-	17.41	Nd
16.81	Unknown	-	5.50	Nd
17.19	Unknown	-	5.80	13.20
21.48	Caffeine	194	31.42	27.99
22.44	Palmitic Acid	256.42	13.42	15.15
24.09	Linoleic acid	280.4	5.44	Nd
24.15	(Z,Z)-9,12-Octadecadienoic acid	280.4	Nd	12.14
24.15	(Z,Z,Z)-9,12,15-Octadecatrienoic-1-ol	264.4	8.31	Nd
24.32	Stearic acid	284.5	3.14	4.39
27.65	Unknown	-	0.63	Nd
27.87	Unknown	-	2.64	Nd
29.88	Unknown	-	0.65	Nd
31.28	Unknown	-	1.81	Nd
34.17	Unknown	-	1.33	Nd

minutes in the stored pulp (Table 4). In contrast to husks, Gayo Arabica coffee pulps had various compounds as detected by GC-MS (Table 5).

The profiles and concentration of secondary metabolites in coffee depend on environmental factors, geographic origin, coffee genetics and variabilities, ripening stages, and pre- and postharvesting processing. [24]. Roasting affects the volatile compounds and aroma of coffee. [1, 13, 24–28]. Some of the volatile compounds identified in roasted coffee are aldehyde, ketones, ester, alcohols, furan, and phenolic volatiles. Drying also alters the compound contents of coffee [1, 24–26]. High temperature decreases the content of aldehyde, ester, acetic, and carboxylic compounds in coffee beans. [12, 13, 24, 26, 29, 30]. Caffeine, or 1,3,7-trimethyl-3,7-dihydro-1H-purine-2,6-dion, is a methylxanthine alkaloid with a neuroprotective agent [14, 16, 18, 31–33]. Caffeine has been detected in coffee beans at high concentrations. However, some coffee bean processing techniques easily degrade caffeine [27, 33].

Coffee compounds promote some biological activities for human health. Chlorogenic acid, caffeine, and quinic acid exhibit anti-inflammatory activity through cyclooxygenase-2 inhibition [15, 34–37]. Caffeic acid stimulates PPAR activities and suppresses ACE2 activities [37, 38]. In addition, chlorogenic acid decreases blood tension [36].

(Z,Z)-9,12 Octadecadienoic acid has antioxidant activity [39], and acetic compounds, such as palmitic acid, exhibit anti-inflammatory and potential anticancer functions [40]. The concentrations of linoleic and palmitic acid in arabica coffee husk and pulp are high at approximately 48%–57% and 26%–28%, respectively [41]. 5-Hydroxymethylfurfural, which is found in coffee, shows antioxidant, antibacterial, and antiproliferative activities [42]. In addition, n-hexadecanoic acid promotes antioxidant activity [29, 43].

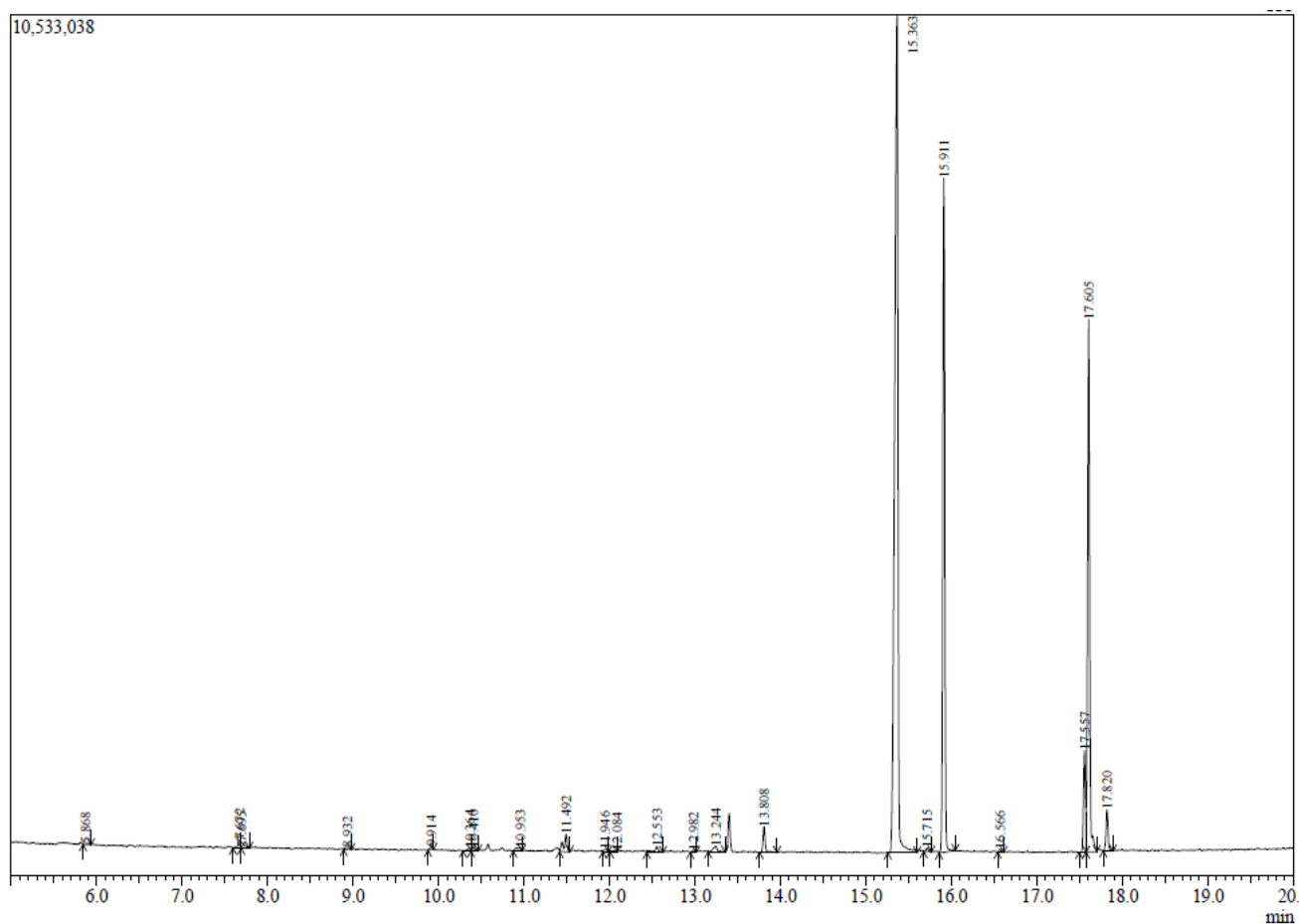


Figure 3. Total Ion Chromatogram of the Fresh Pulp of Gayo Arabica Coffee

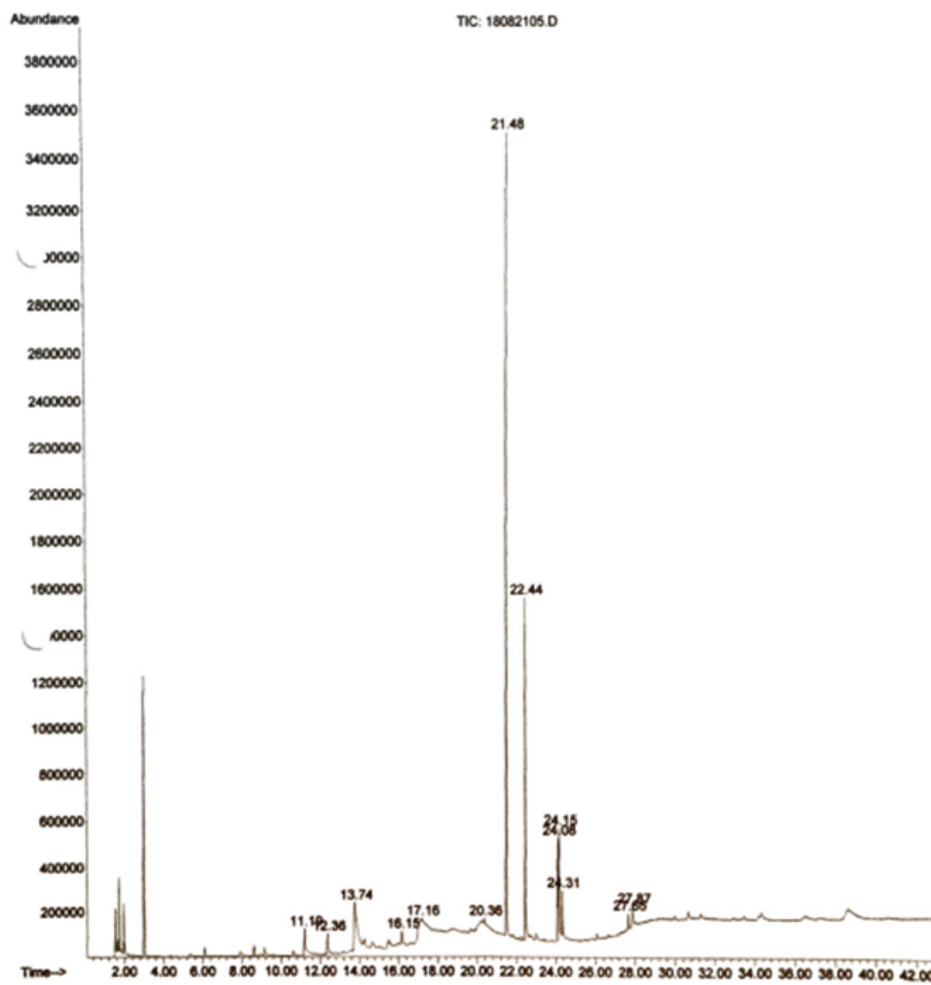


Figure 4. Total Ion Chromatogram of the Long-term Stored Pulp of Gayo Arabica Coffee

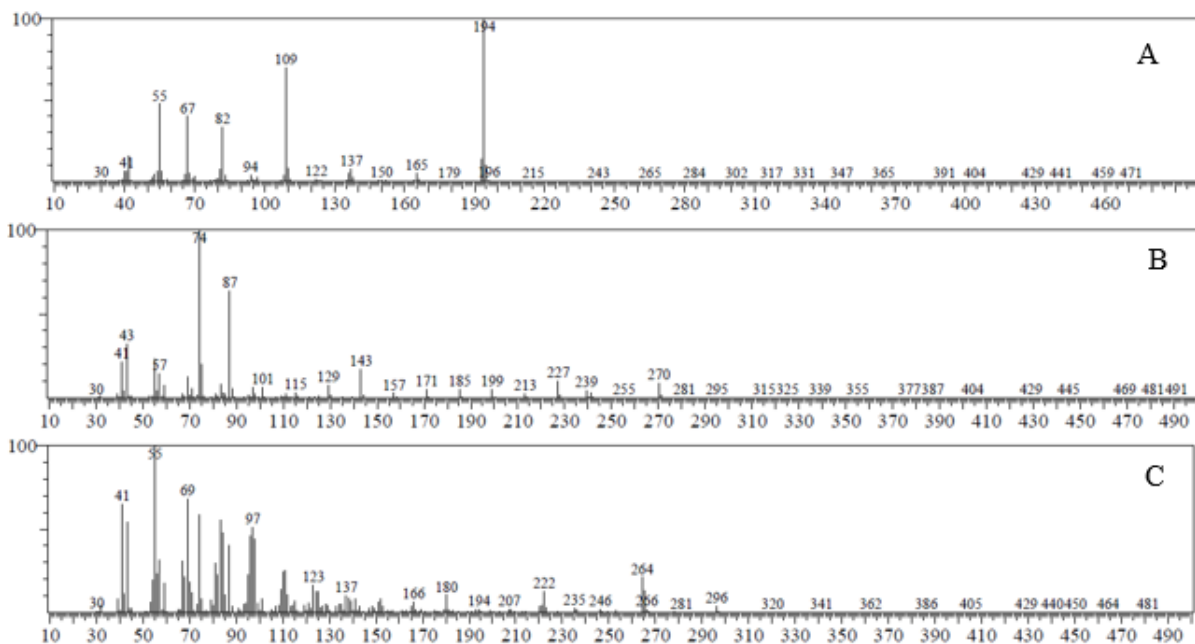


Figure 5. Mass Spectra Detected as Caffeine (A), Hexadecenoic Acid (B), and Elaidic Acid (C) in Gayo Arabica Coffee Fresh Pulp

Table 4. Mass Selective Detector (MSD) of the Long-term Stored Pulp of Gayo Arabica Coffee

Peak	Retention Time (Minute)	First Scan	Max Scan	Last Scan	PK TY	Peak height	Corr. Area	Corr. % max	% of total
1	11.188	1926	1949	1980	BB	115140	5125957	7.63%	2.865%
2	12.361	2101	2145	2179	BB	86635	3572047	5.32%	1.997%
3	13.740	2355	2386	2460	BV	200547	20505465	30.52%	11.462%
4	16.149	2753	2807	2843	BB 4	62127	3903332	5.81%	2.182%
5	17.157	2924	2983	3030	BB 8	66547	12614960	18.78%	7.051%
6	20.361	3441	3543	3576	BB 8	40836	7738156	11.52%	4.325%
7	21.482	3703	3739	3781	BB	3412235	67181158	100.00%	37.552%
8	22.438	3885	3906	3940	BV	1424703	26738585	39.80%	14.946%
9	24.080	4167	4193	4198	BV	428551	7561171	11.25%	4.226%
10	24.143	4198	4204	4227	VV	479638	13319097	19.83%	7.445%
11	24.315	4227	4234	4267	VB	211391	5358206	7.98%	2.995%
12	27.645	4711	4816	4828	BV 8	66694	1365740	2.03%	0.763%
13	27.868	4828	4855	4885	PB 3	87628	3918616	5.83%	2.190%

Table 5. Secondary Metabolite in the Fresh and Long-term Stored Pulps of Gayo Arabica Coffee

Retention Time (Minutes)	Compound	Molecular Weight	% Area of Fresh pulp	% Area of Long-Term Stored Pulp
5.868	2-Pyridinepropanoic acid, .alpha.-methyl-.b	207	0.13	Nd
7.672	5-Hydroxymethylfurfural	126.11	0.41	Nd
7.695	2,4(1H,3H)-Pyrimidinedione, 5-nitro-	306.23	0.34	Nd
8.932	Decanoic acid, methyl ester (CAS) Methyl	186.3	0.18	Nd
9.914	2-Cyclohexene-1-one, 3-(1,3-butadienyl)-2,4	206.33	0.15	Nd
10.364	Carbamic acid, Hydrazinecarboxamide	75	0.18	Nd
10.410	Stearic acid hydrazide	298.5	0.22	Nd
10.953	2H-2,4a-Ethanonaphthalene, 1,3,4,5,6,7-hex	204	0.32	Nd
11.492	Dodecanoic acid, methyl ester (CAS) Methyl	214.34	1.01	Nd
11.946	8'-Apo.beta.-caroten-8'-al	416	0.13	Nd
12.084	3,6-DIHYDRO-3-METHYL-2H-PYRAN-2	112	0.10	Nd
12.553	.gama.-eudesmol	222.37	0.41	Nd
12.982	Ethanedioic acid, dihydrazide (CAS) Oxaly	118.10	0.12	Nd
13.244	.alpha.-Eudesmol, 2-Naphthalenemethanol	222.37	0.40	Nd
13.808	Tetradecanoic acid, methyl ester (CAS) Met	242.40	1.01	Nd
15.363	Caffeine	194	47.66	37.55
15.715	trans-13-Octadecenoic acid, methyl ester	296	0.16	Nd
15.911	Hexadecanoic acid, methyl ester (CAS) Me	270	22.86	Nd
16.566	Malonic acid, dihydrazide	132.12	0.12	Nd
17.557	Linoleic acid, 9,12-Octadecadienoic acid	280.4	3.56	Nd
17.605	Elaidic acid, 9-Octadecenoic acid, methyl e	296.5	19.05	Nd
17.820	Heptadecanoic acid, 16-methyl-, methyl ester	284.5	1.48	Nd
11.19	Unknown	-	Nd	2.87
12.36	Unknown	-	Nd	2.00
13.74	HMF	126.11	Nd	11.46
16.15	Unknown	-	Nd	2.18
17.16	Unknown	-	Nd	7.05
20.36	Unknown	-	Nd	4.33
21.48	Caffeine	194	Nd	37.55
22.44	Palmitic acid	256.42	Nd	14.95
24.08	Linoleic acid	280.4	Nd	4.23
24.15	(Z,Z,Z)-9,12,5-Octadecatrienoic-1-ol	280.4	Nd	7.45
24.31	Stearic acid	284.5	Nd	3.00
27.65	Unknown	-	Nd	0.76
27.87	Unknown	-	Nd	2.19

Nd = not detected

Conclusions

Metabolic investigation using GC-MS revealed that the postharvesting process impacts the secondary metabolites of coffee pulp and husk. Oxidation during storage possibly changes the detected metabolites. Major differences in several acetic acid and ester compounds are noted between the fresh and long-term stored byproducts of Gayo Arabica coffee. In particular, some of these compounds arise or disappear during storage, such as HMF, (Z,Z)-9,12-octadecadienoic acid, and (Z,Z,Z)-9,12,15-octadecatrienoic-1-ol.

Acknowledgments

The author would like to express heartfelt gratitude to Prof. Dr. Kartini Hasballah, MS, Apt, Dr. M. Yulianto Listiawan, Sp.KK(K), FINSDV, FAADV, and Dr. Sofia, M. Sc, for their constant support and direction in writing this paper and conducting this research. Best regards to the faculty of Chemical Engineering and the Mathematic & Natural Sciences Laboratory at Syiah Kuala University for their assistance in providing the GC-MS instrument and extracting the coffee husk and pulp. Thanks to BPSDM (Badan Pengembangan Sumber Daya Manusia) Aceh for the support in this research.

References

- [1] Yeretziyan, C., Opitz, S., Smrke, S., *et al.* 2019. CHAPTER 33. Coffee Volatile and Aroma Compounds—From the Green 200 Bean to the Cup. In Farah, A. (eds.), *Coffee: Production, Quality and Chemistry*. Royal Society of Chemistry. UK. pp. 726–770.
- [2] Lestari, W., Hasballah, K., Listiawan, M.Y., *et al.* 2022. Identification of antioxidant components of gayo arabica coffee cascara using the GC-MS method. *IOP Conf. Ser. Earth Environ. Sci.* 956: 012011, <https://doi.org/10.1088/1755-1315/956/1/012011>.
- [3] Sinaga, H., Hilman, A. 2022. Analysis of cupping, colour and caffeine content of gayo arabica coffee. *IOP Conf. Ser. Earth Environ. Sci.* 977: 012083. <https://doi.org/10.1088/1755-1315/977/1/012083>.
- [4] Syakir, M., Surmaini, E. 2017. Perubahan iklim dalam konteks sistem produksi dan pengembangan kopi di Indonesia. *Jurnal Penelitian dan Pengembangan Pertanian.* 36(2): 77–90, <https://doi.org/http://dx.doi.org/10.21082/jp3.v36n2.2017.p77-90>.
- [5] Badan Perencanaan Pembangunan Daerah Kabupaten Bener Meriah. 2019. Profil Kabupaten Bener Meriah 2019. BAPPEDA Kabupaten Bener Meriah, Aceh.
- [6] Hameed, A., Hussain, S.A., Suleria, H. 2018. “Coffee bean-related” agroecological factors affecting the coffee. In Merillon, J.-M., Ramawat, K.G. (eds.), *Co-Evolution of Secondary Metabolites*. Springer Cham. Denmark. pp. 1–67.
- [7] Nwokeji, P.A., Enodiana, O.I., Ezenweani, R., *et al.* 2016. The chemistry of natural product: Plant secondary metabolites. *Nat. Prod.* 4(8): 1–8.
- [8] Velu, G., Palanichamy, V., Rajan, A. 2018. Phytochemical and Pharmacological Importance of Plant Secondary Metabolites in Modern Medicine. In Roopan, S.M., Madhumitha, G. (eds.), *Bioorganic Phase in Natural Food: An Overview*. Springer Cham. Denmark. pp. 135–156.
- [9] Komaria, N., Suratno, Prihatin, J., *et al.* 2020. An analysis of innovation on the utilization of cascara by coffee farmers. *J. Phys. Conf. Ser.* 1563: 012015, <https://doi.org/10.1088/1742-6596/1563/1/012015>.
- [10] Al-Youssef, H.M., Al-Yousef, H.M., Sawab, A., *et al.* 2017. Pharmacognostic studies on *Coffea Arabica* L. husks: A brilliant source of antioxidant agents. 4(1): 86–92.
- [11] Dimberg, L. 2015. A nutritional analysis on the by-product coffee husk and its potential utilization in food production. *Food Sci. Technol.* 415.
- [12] Klingel, T., Kremer, J.I., Gottstein, V., *et al.* 2020. A review of coffee by-products including leaf, flower, cherry, husk, silver skin, and spent grounds as novel foods within the european union. *Foods.* 9(5): 1–20, <https://doi.org/10.3390/foods9050665>.
- [13] Ameca, G.M., Cerrilla, M.E.O., Córdoba, P.Z., *et al.* 2018. Chemical composition and antioxidant capacity of coffee pulp. *Cienc. Agrotec.* 42(3): 307–313, <https://doi.org/10.1590/1413-70542018423000818>.
- [14] Wasim, S., Kukkar, V., Awad, V.M., *et al.* 2020. Neuroprotective and neurodegenerative aspects of coffee and its active ingredients in view of scientific literature. *Cureus.* 12(8): 5–11, <https://doi.org/10.7759/cureus.9578>.
- [15] Bare, Y., Sari, D.R.T., Rachmad, Y.T., *et al.* 2019. In silico insight the prediction of chlorogenic acid in coffee through Cyclooxygenase-2 (COX2) interaction. *Biogenesis: Jurnal Ilmiah Biologi.* 7(2): 100–105, <https://doi.org/https://doi.org/10.24252/bio.v7i2.9847>.
- [16] Kolahdouzan, M., Hamadeh, M.J. 2017. The neuroprotective effects of caffeine in neurodegenerative diseases. *CNS Neurosci. Ther.* 23(4): 272–290, <https://doi.org/10.1111/cns.12684>.
- [17] Silva, M.R.d., Bragagnolo, F.S., Carneiro, R.L., *et al.* 2019. Metabolite characterization of fifteen by-products of the coffee production chain: From farm to factory. *Food Chem.* 369: 130753, <https://doi.org/10.1016/j.foodchem.2021.130753>.
- [18] Lee, K.E., Bharadwaj, S., Yadava, U., *et al.* 2019. Evaluation of caffeine as inhibitor against collagenase, elastase and tyrosinase using in silico and in vitro approach. *J. Enzyme Inhib. Med. Chem.* 34(1): 927–936, <https://doi.org/10.1080/14756366.2019.1596904>.

- [19] Safaryani, N., Haryanti, S., Hastuti, E.D. 2007. Pengaruh suhu dan lama penyimpanan terhadap penurunan kadar vitamin C brokoli (*Brassica oleracea* L). Buletin Anatomi dan Fisiologi. XV(2): 39–45, <https://doi.org/10.14710/baf.v15i2.2571>.
- [20] Rakić, S., Janković, S., Marčetić, M., *et al.* 2014. The impact of storage on the primary and secondary metabolites, antioxidant activity and digestibility of oat grains (*Avena sativa*). J. Funct. Foods. 7: 373–380, <https://doi.org/10.1016/j.jff.2014.01.022>.
- [21] Hamzahoğlu, A., Gökmen, V. 2018. Investigation and kinetic evaluation of the reactions of hydroxymethylfurfural with amino and thiol groups of amino acids. Food Chem. 240: 354–360, <https://doi.org/10.1016/j.foodchem.2017.07.131>.
- [22] Kumar, S.S., Swapna, T.S., Sabu, A. 2018. Coffee Husk: A Potential Agro-Industrial Residue for Bioprocess. In Singhania, R.R., Agarwal, R.A., Kumar, R.P., *et al.* (eds.), Waste to Wealth. Springer Singapore. Singapore. pp. 97–109.
- [23] Orkusz, A., Wolańska, W., Krajinska, U. 2021. The assessment of changes in the fatty acid profile and dietary indicators depending on the storage conditions of goose meat. Molecules. 26(17): 5122, <https://doi.org/10.3390/molecules26175122>.
- [24] Zhang, K., Cheng, J., Hong, Q., *et al.* 2022. Identification of changes in the volatile compounds of robusta coffee beans during drying based on HS-SPME/GC-MS and E-nose analyses with the aid of chemometrics. Lwt. 161: 113317, <https://doi.org/10.1016/j.lwt.2022.113317>.
- [25] Sunarharum, W.B., Williams, D.J., Smyth, H.E. 2014. Complexity of coffee flavor: A compositional and sensory perspective. Food Res. Int. 62: 315–325, <https://doi.org/10.1016/j.foodres.2014.02.030>.
- [26] Caporaso, N., Whitworth, M.B., Cui, C., *et al.* 2018. Variability of single bean coffee volatile compounds of Arabica and robusta roasted coffees analyzed by SPME-GC-MS. Food Res. Int. 108: 628–640, <https://doi.org/10.1016/j.foodres.2018.03.077>.
- [27] Cangussu, L.B., Melo, J.C., Franca, A.S., *et al.* Chemical characterization of coffee husks, a by-product of *coffea arabica* production. Foods. 10(12): 3125, <https://doi.org/10.3390/foods10123125>.
- [28] Moreno, J., Cozzano, S., Pérez, A.M., *et al.* 2019. Coffee pulp waste as a functional ingredient: Effect on salty cookies quality. J. Food Nutr. Res. 7(9): 632–638, <https://doi.org/10.12691/jfnr-7-9-2>.
- [29] Kavitha, R., Uduman, M.A.M. 2017. Identification of bioactive components and its biological activities of abelmoschus moschatus flower extract. A Gc-Ms Study. IOSR J. Appl. Chem. 10: 19–22.
- [30] Manasa, V., Padmanabhan, A., Appaiah, K.A.A. 2021. Utilization of coffee pulp waste for rapid recovery of pectin and polyphenols for sustainable material recycle. Waste Manag. 120: 762–771, <https://doi.org/10.1016/j.wasman.2020.10.045>.
- [31] Camandola, S., Plick, N., Mattson, M.P. 2019. Impact of coffee and cacao purine metabolites on neuroplasticity and neurodegenerative disease. Neurochem. Res. 44: 214–227, <https://doi.org/10.1007/s11064-018-2492-0>.
- [32] Socala, K., Szopa, A., Serefko, A., *et al.* 2021. Neuroprotective effects of coffee bioactive compounds: A review. Int. J. Mol. Sci. 22(1): 1–64, <https://doi.org/10.3390/ijms22010107>.
- [33] Li, S., He, B., Wang, J., *et al.* 2020. Risks of caffeine residues in the environment: Necessity for a targeted ecopharmacovigilance program. Chemosphere. 243: 125343, <https://doi.org/10.1016/j.chemosphere.2019.125343>.
- [34] Bare, Y., Kuki, A.D., Rophi, A.H., *et al.* 2019. Prediksi asam kuinat sebagai anti-inflamasi terhadap COX-2 secara virtual. Biota Jurnal Ilmiah Ilmu-Ilmu Hayati. 4(3): 124–129, <https://doi.org/10.24002/biot.a.v4i3.2516>.
- [35] Bare, Y., S M., Putra, S.H.J., *et al.* 2020. In-silico approach for the prediction of chlorogenic acid as PPAR- γ activator. Biota Biologi dan Pendidikan Biologi. 13(1): 68–77, <https://doi.org/10.20414/jb.v13i1.197>.
- [36] Bare, Y., Sari, D.R., Rachmad, Y.T., *et al.* 2019. Prediction potential chlorogenic acid as inhibitor ace (In silico study). Bioscience. 3(2): 197–203, <https://doi.org/10.24036/0201932105856-0-00>.
- [37] Bare, Y., Kuki, A.D., Tiring, S.S.N.D., *et al.* 2020. In silico study: Prediction the potential of caffeic acid as ACE inhibitor. El-Hayah Jurnal Biologi. 7(3): 94–98, <https://doi.org/10.18860/elha.v7i3.10053>.
- [38] Bare, Y., Helvina, M., Elizabeth, A., *et al.* 2019. Potensi asam kafeat pada kopi sebagai simultan gen peroxisome *Proliferator-activated receptor gamma* (Ppar- γ 3 277): Studi in Silico. Jurnal Saintek Lahan Kering. 2(2): 52–53, <https://doi.org/10.32938/slk.v2i2.866>.
- [39] Tian, C., Gao, X., Yang, J., *et al.* 2018. Chemical compositions, extraction technology, and antioxidant activity of petroleum ether extract from *Abutilon theophrasti* medic. leaves. Int. J. Food Prop. 21(1): 1789–1799, <https://doi.org/10.1080/10942912.2018.1494198>.
- [40] Korbecki, J., Bajdak-Rusinek, K. 2019. The effect of palmitic acid on inflammatory response in macrophages: an overview of molecular mechanisms. Inflamm. Res. 68: 915–932, <https://doi.org/10.1007/s00011-019-01273-5>.
- [41] Mehari, B., Redi-Abshiro, M., Chandravanshi, B.S., *et al.* 2019. GC-MS profiling of fatty acids in green coffee (*Coffea arabica* L.) beans and chemometric modeling for tracing geographical origins from Ethiopia. J. Sci. Food Agric. 99(8): 3811–3823, <https://doi.org/10.1002/jsfa.9603>.

[42] Yassin, M., Mostafa, A., Al Askar, A. 2021. In Vitro Evaluation of Biological Activities and Phytochemical Analysis of Different Solvent Extracts of *Punica granatum* L. (Pomegranate) Peels. *Plants*. 10(12): 1–18, <https://doi.org/10.3390/plants10122742>.

[43] Mellado, M., Soto, M., Madrid, A., et al. 2019. In vitro antioxidant and antiproliferative effect of the extracts of *Ephedra chilensis* K Presl aerial parts. *BMC Complement. Altern. Med.* 19(53): 1–10, <https://doi.org/10.1186/s12906-019-2462-3>.