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Gunawan Indrayanto Faculty of Pharmacy, Universitas Surabaya, Surabaya 60293, Indonesia, gunawanindrayanto@staff.ubaya.ac.id

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Application of Accuracy and Precision Evaluations Based on the Current United States and Indonesian Pharmacopoeias: A Critical Review

Gunawan Indrayanto*

Faculty of Pharmacy, Universitas Surabaya, Surabaya 60293, Indonesia

*E-mail: gunawanindrayanto@staff.ubaya.ac.id

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Abstract

New methods for the evaluation of accuracy and precision are mentioned in the latest edition of the United States Pharmacopoeia (USP), whereas other validation parameters, that is, selectivity, linearity, range, and robustness, remained relatively unchanged. In obtaining reliable data from any chemical/pharmaceutical analysis, the analytical procedure must be validated or verified in accordance with the latest edition of the pharmacopoeia. Some review articles on the general validation methods have been published by the author. This present review will focus on the implementation and discussion of the accuracy and precision evaluation based on the current USP and Indonesian pharmacopoeia. Some examples of the calculation of several accuracy and precision method of determinations are also discussed.

Keywords: accuracy, intermediate precision, repeatability, validation method

Introduction

Based on the general chapter of United States Pharmacopoeia (USP)-National Formulary (US44-NF39) <1225> [1], the accuracy of the analytical procedure can be defined as the closeness of test results obtained by that procedure to the true value. The accuracy of an analytical procedure should be established across its range, whereas precision is the degree of agreement among individual test results when the procedure is applied repeatedly to multiple samplings of a homogeneous sample. The definition of accuracy and precision in the general chapter USP-NF39 <1225> is almost similar to their definition in British Pharmacopeia (BP) 2022, Supplementary III F [2], International Council for Harmonization (ICH) Q2R1 [3], and Indonesian Pharmacopoeia (FI) VI <1381> [4]. The ICH of Technical Requirements for Pharmaceuticals for Human Use O2R1 uses the term trueness instead of accuracy. According to International Organization for Standardization (ISO) 5725-2:2019 [5], the term accuracy indicates the combination of the term trueness and precision. General chapter USP44-NF39 <1225> and FI VI <1381> [1, 4] describe general validation methods of analytical procedures and their acceptance criteria; the method performance is determined using the parameters accuracy, precision, specificity, detection limit, quantification limit, linearity, and range. General chapter USP44-NF39 <1210> [6] describes the utilization of statistical approaches in procedure validation as described in chapter <1225> [1]. This chapter focuses on

establishing analytical performance characteristics of accuracy, precision, and detection limit. Accuracy can only be evaluated if a true or accepted reference value is available. Accuracy of an analytical procedure expresses the closeness of agreement between $\boldsymbol{\tau}$ (true or nominal value) and Y (measured value); the closeness is expressed as the average of $(Y - \tau)$. General chapter USP44-NF39 <1010> [7] provides a basic statistical approach for decision-making and the methods for comparison of two analytical procedures; comparing two analytical procedures (e.g., new- and validated-method) is necessary to determine whether the difference of the accuracy and precision are less than the amount described in the analytical target profile (ATP). General Notices and Requirements Section 6.30 of the USP [8] describes the need to produce comparable results from the proposed/alternative to the compendial method. This comparative test described in general chapter USP 44-NF 39 < 1010 > can be also applied to the method validation for transferring analytical procedures in different laboratories [7]. The performance characteristic of accuracy and precision, which should be evaluated and specified in the method validation, must meet the acceptance criteria described by the ATP of the proposed/new method [9]. Review articles on the validation of analytical methods and their applications in pharmaceutical analysis (including herbal drugs) have been published by the author in 2005 [10], 2012 [11], 2018 [12, 13], and 2022 [14, 15]. In the last 4 years, new methods for the evaluation of accuracy and precision are mentioned in the latest edition of the US, whereas other

validation parameters, that is, selectivity, linearity, range, and robustness, remained relatively unchanged. The analytical method must be validated/verified based on the latest edition of the pharmacopoeia or official guidelines to obtain reliable data from any chemical analysis carried out by QC or research laboratories; if a new edition of the pharmacopoeia has been released, then the old version will automatically no longer be applied.

This review will describe and focus on implementing accuracy and precision evaluation in accordance with the current USP44-NF39 [1, 6, 7, 9] and FI VI [4]. It also aims to gain a comprehensive understanding of the new methods for determining accuracy and precision based on those current Pharmacopoeias. Related official guidelines, publications, and some similarities from previous editions of the USP-NF will be described in this review because of their important role in current evaluation methods. Some examples of calculations and determinations of accuracy and precision are also described. Given their comprehensive understanding of accuracy and precision evaluation methods, the pharmacists in QC laboratories and/or researchers can select the best method for a particular application in their work.

Assessment of Accuracy and Precision

Separated Evaluation of Accuracy and Precision: According to chapters USP44-NF39 <1225> [1] and FI VI <`1381>[4]. Accuracy can be determined by three categories: (1) Drug substance: accuracy is determined by applying an analytical procedure to an analyte of known purity (e.g., a certified reference standard) or by comparing with a well-characterized procedure, where the accuracy has been stated or defined. (2) Drug in product: accuracy can be determined by applying an analytical procedure to a synthetic mixture of the drug product components to which a certain amount of analyte is added within the range of the procedure. If a sample with all components of the drug product can be obtained, that is, active pharmaceutical ingredient (API) and excipients, then this can be done by adding a known amount of analyte to the drug product (spiking) or by comparing with a well-characterized procedure, where the accuracy has been stated or defined. (3) Impurities: accuracy should be assessed on a sample (of drug substance or drug product) that is spiked with a known amount of impurity.

Accuracy should be assessed using a minimum of nine determinations over a minimum of three concentration levels, covering the specified range (i.e., three concentrations and replicates of each concentration). The accuracy can be evaluated as follows: (1) Determine the percent of recovery (%R) across the range of the assay. (3) Evaluate the linearity of the relationship between the estimated and actual concentrations. The statistically

preferred criterion indicates that the confidence interval (CI) for the slope is contained within an interval of about 1.0 or alternatively, that is, the slope is close to 1.0.

% R can be calculated using Equation 1 or 2 (standard addition method).

or

$$\%R = \frac{Y_f}{Y_c} \times 100 \tag{1}$$

$$\% R = \frac{Y_f - Y_u}{C_a} \times 100 \tag{2}$$

where Y_c is the actual or true concentration, Y_f is the measured/estimated concentration, Y_u is the original concentration before standard addition, and C_a is the added concentration of the analyte. The acceptance criteria of certain drug substance or product are described by their USP-NF and FI monographs, or it can be referred to Tables 1 and 2 (if the monograph is not yet available). The acceptance criteria depend on analyte concentrations in samples or the instrument used, which should be stated in the ATP of the proposed method. Researchers must decide whether the acceptance criteria to be specified in the ATP are based on the concentration of API in the sample and/or the instrument used. Food and Drug Administration Office of Regulatory Affairs Laboratory Manual II [16] describes that the general acceptance criteria of accuracy for human drug analytical methods is at least 80%-120% (for assay) and 70%-130% (for content uniformity) of the expected content.

USP <1225> [1] and FI <1381> [4] do not describe the method for the evaluation of the linearity relationship of the recovery curve between Y_f and Y_c (Equation 3). Funk *et al.* [28] described the equations used to determine the CI of the slope (b) and intercept (a) (Equations 4 and 5) of the recovery curve. The CI value must include the value of 1 (slope) and 0 (intercept); if it does not include the respected values, then one may assume a constant and/or proportional systematic error of the proposed method.

$$Y_f = \mathbf{a} + \mathbf{b} \cdot Y_c \tag{3}$$

$$CI_{b} = b \pm \frac{t \cdot S_{y}}{\sqrt{Q_{XX}}}$$
(4)

$$S_{y} = \sqrt{\frac{\sum[Y_{if} - (a+b.\ Y_{ic})^{2}}{n-2}}; Q_{XX} = \sum Y_{ic}^{2} - \frac{1}{n} (\sum Y_{ic})^{2}$$
$$CI_{a} = a \ \pm t. S_{y} \sqrt{(\frac{1}{n} + \frac{\bar{Y}_{c}^{2}}{Q_{XX}})}$$
(5)

i (1, 2 ..., n) indicates different concentration levels, and *t* is the student-*t*-factor based on the degree of freedom of n - 2 (p = 0.05). Recently, ICH Q14 described that the slope of the recovery curve between Y_f and Y_c should be within 0.8 to 1.25 (for p = 0.05) [29].

Concentration (C)	%Recovery Limit	% Bias Limit (λ)**	Sr***	S _R ***
100%	98-101	-2 to +1	1	2
10%	95-102	-5 to +2	1.5	3
1%	92-105	-8 to +5	2	4
0.1%	90–108	-10 to $+8$	3	6
0.01%	85-110	15 to +10	4	8
10 µg/g (ppm)	80-115	-20 to 15	6	11
1 ppm	75–120	-25 to +20	8	16
10 µg/kg (ppb)	70–125	-30 to +25	15	32

Table 1. Acceptance Criteria of Accuracy and Precision*

*Data from Ref. [17]; ** calculated from % Recovery; ***Limit of S_r (repeatability) and S_R (intermediate precision) are calculated as RSD/CV. The acceptance criteria of the cumulative variance in laboratories (S_R) can be estimated as $1.6 \times S_r$; S_r and S_R can also be estimated using equation $2C^{-0.15}$ (C is the mass fraction) [17].

Table 2. Acceptance Criteria of the Accuracy and Precision of Various Instruments

	Accuracy (%)		Precision (% RSD)			Ref.				
Instrument	DS	DS DP Imp.	Imn	Repeatability			Intermediate precision			
	DS	DP	Imp.	DS	DP	Ι	DS	DP	Imp.	
MIR	98–102	95-105	70–150	1	2	20	1	3	25	18
NIR	98-102	95–05	70–150	1	2	20	1	3	25	19
UV/VIS	98-102	95-105	80-120	1	2	15-20	1.5	3	15-25	20
AAS	95-105	95-105	70–150	5	5	20	8	8	25	21
ICP	Na	95-105	70–150	5	5	20	8	8	25	22
Fluorescence Spectroscopy	98–102	95–105	80–120	1	2	20	1	3	25	23
Raman	98-102	95-105	70–150	1	2	20	1	3	25	24
NMR	98-102	95-105	80-120	1	2	20	1	3	25	25
MS	Na	80-120	50-120	Na	Na	Na	Na	Na	Na	26
Chromatography*	Na	Na	Na	Na	Na	Na	Na	Na	Na	27

DS: drug substance, DP: drug product, Imp: impurities, Na: not available. *The acceptance criteria can be referred to the detector of the chromatographic system.

In general, the precision of an analytical procedure is expressed as the standard deviation (SD), relative standard deviation (RSD), or coefficient of variation (CV) of a series of measurements [1, 4]. Three levels of precision determination include repeatability, intermediate precision (ruggedness), and reproducibility [30]. Repeatability refers to the use of an analytical procedure within a laboratory over a short period of time using the same analyst with the same equipment. Intermediate precision (also known as ruggedness) expresses withinlaboratory variation, as on different days, or with different analysts or equipment within the same laboratory. Reproducibility refers to the use of an analytical procedure in different laboratories, as in a collaborative study [1, 4].

Precision is determined by testing a sufficiently homogenous sample aliquot and expressed as SD or

RSD. Sample analysis must be performed through a complete analytical procedure from sample preparation to final test result. Repeatability must be assessed using a minimum of nine determinations covering the specified range of the procedure (*i.e.*, three concentration levels and three replicates of each concentration) or using a minimum of six determinations at 100% of the test concentration [1, 4]. For analysis using a single run, SD can be calculated using Equation 6.

$$SD = \sqrt{\frac{\sum_{i=1}^{n} (Y_i - \bar{Y})^2}{n-1}}; RSD = CV = \frac{SD}{Mean} \times 100\%$$
(6)

where *Yi* is the individual value, and \overline{Y} is the sample mean. Current Pharmacopoeias [1, 4] do not include equation for calculating the intermediate precision (S_R); S_R can be calculated using Equation 7 as described by Ermer [30] and discussed in a previous review

article [11]. Based on Ref. [30], S_R can also indicate reproducibility.

$$\mathbf{S}_{\mathrm{R}} = \sqrt{S_r^2 + S_B^2} \tag{7}$$

where S_r is the repeatability, and S_B is between condition variance; S_B , S_r , and SD of the mean (SD_m) can be determined using ANOVA (Equations 8 and 9), which were described in the previous edition of the USP41-NF36, 2018 [31]. Detailed discussions can be referred to author's previous publication [11], but the equations were not mentioned in the current edition of the USP [7].

$$S_{\rm B} = \frac{MS_{betwwen} - MS_{withn}}{replicates \ per \ run} \tag{8}$$

$$\mathbf{S}_{\mathrm{r}} = \sqrt{MS_{withn}} \tag{9}$$

The previous edition of the USP [31] described the variance of the mean (V_m) for a test involving different combinations of runs and number of replicates per run using (Equation 10):

$$\mathbf{V}_{\mathrm{m}} = \frac{S_B}{m} + \frac{S_T}{m.n} \tag{10}$$

$$SD_m = \sqrt{V_m}$$

where m is the number of runs, and n is the number of replications for each of run. Equation 10 is no longer described in the current edition of the USP-NF [7]. For in-house validation of a new analytical procedure, the author recommends determining S_R instead of S_r for precision evaluation. The precision acceptance criteria of certain drugs and preparations have been described by the monographs of the pharmacopoeia or Tables 1 and 2. FDA, ORA Laboratory Manual II [16] describes the general acceptance criteria for the precision of human drug analytical methods, which are <3% (for drug products) and <2% (for API).

General chapter of USP44-NF39 <1210>[6]. Accuracy and precision can be evaluated using CI of bias (CI_B) and CI of SD (CI_{SD}).

$$CI_{B} = (\overline{Y} - \tau) \pm t_{1-\alpha,n-1} \times \frac{SD}{\sqrt{n}}$$
(11)

$$CI_{SD} = SD \sqrt{\frac{n-1}{x_{\alpha:n-1}^2}}$$
(12)

$$SD = \sqrt{\frac{\sum_{i=1}^{n} (Y_i - \bar{Y})}{n-1}}$$

where τ is the true or nominal value, bias (B) is $(\overline{Y} - \tau)$, n is the number of reportable value, $t_{1-\alpha,n-1}$ is the percentile of central t-distribution with area $1 - \alpha$ to the left and (n - 1) degrees of freedom, $X_{\alpha:n-1}^2$ is a percentile of a

central chi-squared distribution with area α to the left, and (n-1) degrees of freedom.

The acceptance criteria of CI_B and CI_D can be referred to Tables 1 and 2. If λ is the maximum bias acceptance limit, then CI_B must be between $-\lambda$ and $+\lambda$, and CI_{SD} should be less than the acceptance values. If the nominal content of the API is 0.1% (Table 1), then CI_B should be within -10% to +8%, and CI_{SD} must be <3% (repeatability) or <6% (reproducibility). If the researcher using NIR (Table 2) limit of $\%\lambda$ ranging from -2% to +2% (DS), from -5% to +5% (DP), or from -30% to +50% (impurities). The researcher must determine whether to use acceptance criteria based on nominal concentration or the instrument used must be described in ATP.

Combined evaluation of accuracy and precision. Using separated accuracy and precision as discussed in section separated evaluation of accuracy and precision, some data of the individual results of Y_i or %R are not included in the required acceptance criteria (Tables 1 and 2). Several methods used for evaluating accuracy and precision simultaneously have been proposed to ensure that all Y_i or %R can meet the requirement of the acceptance criteria; discussion of this method has been described by a previous review [12].

The general chapter USP44-NF39 <1210> [6] describes the methods for the simultaneous assessment of accuracy and precision. Y_i or %R should fall between $(-\lambda + \tau)$ and $(\lambda + \tau)$, indicating that $\overline{Y} \pm$ prediction interval (PI)/tolerance interval (TI) must be included in the permitted specification range (Tables 1 and 2).

Mean
$$\pm$$
 PI = $\overline{Y} \pm t_{(1+P)/2,n-1} \times \text{SD} \sqrt{1 + \frac{1}{n}}$ (13)

$$Mean \pm TI = \overline{Y} \pm K. SD \tag{14}$$

$$\mathbf{K} = \sqrt{\frac{Z_{1+P}^2 X(n-1)}{Z_{\alpha,n-1}^2}} x(1+\frac{1}{n})$$

where $t_{(1+P)/2, n-1}$ is the percentile of a central t-distribution with area (1 + P)/2 to the left and (n - 1) degrees of freedom, $Z_{1+P/2}^2$ is the square of the standard normal percentile with area (1 + P)/2 to the left, and $X_{\alpha,n-1}^2$ is the chi-squared percentile with area α to the left and (n - 1)degrees of freedom.

Japan Pharmacopeia 17th edition [32] used $\overline{Y} \pm CI$ to assess accuracy; CI must be calculated using intermediate precision or reproducibility. $\overline{Y} \pm CI$ should be included in the range of the acceptance criteria (Tables 1 and 2). The application of CI (p = 0.05) in evaluating recovery was also recommended by the new version of ICH Q2 (R2) [33]. In evaluating the percent of the data ($\overline{Y} \pm \text{CI/PI/TI}$) included in the required specification range (P%)_{inside} can be calculated using Excel [34]:

$$P (\%) inside = NORM.DIST(SL_{upper}; \mu;SD;TRUE) - NORM.DIST(SL_{lower};\mu;SD;TRUE)$$
(15)

where SL_{upper, lower} is the upper or lower specification limit of the ATP. These discussions show that every results of the pharmaceuticals analysis at QC laboratory ($\overline{Y} \pm$ CI/PI/TI) should be included in the acceptance criteria (Tables 1 or 2), or P (%)_{inside} should be close to 100%. The author recommends applying this combination methods rather than the separate methods, which are discussed in section separated evaluation of accuracy and precision.

Selection of SD and the methods of evaluations. As previously discussed, SD can be calculated as Sr, SR, and SD_m. For analysis, which is performed using several series of measurements (runs), the values of S_R would be > S_r and > SD_m as shown by a previous work [35]. SD can be applied for a single run of analysis. Calculations of accuracy and precision using Equations 11-15 require SD value; the results of calculation of the mean \pm PI/TI (Equations 13 and 14) yield broadest expected ranges, if S_R is used as SD; this shows that the application of S_R as SD is the best choice. Our work [35] showed that the calculation of \overline{WR} and precision using separate evaluations (Equations 1, 2, 4, 5, 11, and 12) and the combination evaluation (Equation 13) met the acceptance criteria of the ATP, but calculation using Equation 14 showed that some data were out of specification (OOS). Therefore, the combination evaluation using TI (Equation 14) is recommended for the evaluation of accuracy and precision because it could yield a broader calculated expected range. If Equation 14 is used, and the specification range of the ATP is achieved, then using other equations will also meet the ATP. Detailed discussion regarding this matter has been described in previous publication by the author [12].

Assessment of the Accuracy and Precision Through Comparison

Principle of evaluation. Based on USP44-NF39 <1225> [1], accuracy and precision of the proposed/new (N) method can be validated by comparing with the old (O) validated method. The detailed comparison methods have been described by the current USP [7], which will be discussed below.

Comparing the accuracy of two procedures, the absolute value of the true difference in means (μ_D) can be calculated as follows.

$$|\mu_D| = |\mu_N| - |\mu_O|$$
(16)

 $|\mu_D|$ must be less than the required value (d). For precision, the SD ratio of the new procedure to the old procedure must be less than a certain required value (k).

$$\frac{SD_N}{SD_O} \text{ must be } < k \tag{17}$$

If an old procedure has μ_0 of 100 unit, then upper and lower specification limits are 104 and 96, respectively, CV₀ is 0.16, and CV_{lot variance} is 0.64. The probability of OOS for several values of d and k can be estimated as follows:

$$\operatorname{Prob}\left(\operatorname{OOS}\right) = 1 - \emptyset\left(\frac{104 - (100 + d)}{\sqrt{0.64 + k^2 x 0.16}}\right) + \emptyset\left(\frac{96 - (100 + d)}{\sqrt{0.64 + k^2 x 0.16}}\right) (18)$$

where \emptyset represents the cumulative probability function of the normal standard distribution. Table 3 describes the expected OOS of the new procedure for various d and k values. If d = 1 and k = 2 is selected, then the OOS of the new procedure will be 0.14%; if d = 2 and k = 1, then OOS will be 1.27%. OOS of an analytical procedure can be calculated using Equation 19 [36]:

Prob (OOS) = 2.
$$\emptyset$$
 (-3. Cp) (19)

where \emptyset represents the normal standard cumulative distribution function (Z value), and the process capability (C_P) can be estimated using Equation 20 [12].

$$Cp = \frac{SL_{upper} - SL_{lower}}{6 SD}$$
(20)

OOS can also be estimated by 100% minus P (%) $_{inside}$ (Section assessment of the accuracy and precision through comparison).

Based on BP 2022 [2], the accuracy evaluation of two procedures could be evaluated using cross-correlation coefficient r (Pearson Product–Moment Correlation). If N is the proposed procedure, then O is the validated procedure. r can be calculated using Equation 21 [37]:

$$r = \frac{n(\Sigma Y_N, Y_O) - (\Sigma Y_N) \cdot (\Sigma Y_O)}{\sqrt{[n(\Sigma Y_N^2) - (\Sigma Y_N)^2][n(Y_O^2) - (\Sigma Y_O)^2]}}$$
(21)

Based on ICH M10 [38], the assessment of the accuracy by comparison can be evaluated using the concordance correlation coefficient (CCC) as described by Equation 22 [39].

Table 3. OOS percentage (%) of the New Procedure*

k = 1	k = 1.5	k = 2
0.001	0.01	0.04
0.04	0.14	0.40
1.27	2.28	3.85
	0.001 0.04	0.001 0.01 0.04 0.14

*Modified from Ref. [7]

$$CCC = \frac{2r.SD_N.SD_0}{(Y_N - Y_0)^2 + SD_N^2 + SD_0^2}$$
(22)

CCC of 0.90, 0.90–0.95, 0.95–0.99, and >0.99 indicates poor, moderate, substantial, and perfect, respectively [40].

Evaluating the accuracy and precision of the proposed procedure by comparing with a validated procedure using a significance test with a certain level of p is not recommended [41, 42].

Comparison using homogenous test materials [7]. The number of replications of the new procedure (n_N) and the old procedure (n_O) can be calculated using Equation 19.

$$n_{\rm N} = n_{\rm O} = 2 \times \left(\frac{(Z_{1-\alpha} + Z_{1-\beta)x\,SD_0}}{d - |\mu_D|}\right)^2 + 1 \tag{21}$$

where $Z_{1-\alpha}$ and $Z_{1-\beta}$ are the standard normal percentiles with area $1-\alpha$ and $1-\beta$, respectively, to the left. The Type I error rate is α , and the Type II error rate is β . Table 4 presents the power $(1 - \beta)$ for sample size combination with d = 1, k = 2, $\alpha = 0.05$, and $SD_N = SD_O = 0.4$ (for μ of 100 unit). Samples size should be 15 to obtain around 0.80 power of the new procedure. The accuracy of the two procedures is tested by calculating CI of μ_D , which should be fulfilled $-d < \mu_D < +d$, and CI_{SD} must be <k.

$$CI_{\mu_D} = \bar{Y}_N - \bar{Y}_O \pm t_{1-\alpha:df} \sqrt{\frac{SD_N^2}{n_N} + \frac{SD_O^2}{n_O}}$$
(22)

$$df = \frac{\left(\frac{SD_N^2 + SD_0^2}{n_N + n_0}\right)^2}{\frac{SD_N^4}{n_N^2(n_N - 1)} + \frac{SD_0^4}{n_0^2(n_0 - 1)}}$$
$$CI_{SD} = \frac{SD_N}{SD_0} \sqrt{\frac{1}{F_{\propto, n_N - 1, n_0 - 1}}}$$
(23)

where $t_{1-\alpha:df}$ is a quantile from a central *t*-distribution with area $1 - \alpha$ to the left and degrees of freedom (*df*), and F α , n_{N-1} , n_{O-1} is the *F*-quantile with area α to the left and degrees of freedom n_{N-1} and n_{O-1} .

Table 4. Power Values for Noninferiority Test ($\alpha = 0.05$)*

0	Homogenous Test Materials		ion Across Material
n	Power	n	Power
8	0.528	8	0.391
14	0.781	17	0.775
15	0.808	18	0.803
19	0.890	22	0.885
20	0.904	23	0.901

*Data from Ref. [7]; α : Type I error, Power = $1 - \beta$ (β is Type II error), d = 1, k = 2, and SD₁ = SD₂ = 0.4 for \overline{Y}_2 of 100 units; n = required replications.

Comparison of two procedures using nonhomogenous test materials (variation across test samples) [7]. For this paired design using nonhomogenous samples (different lots and manufactures), the number of replications can be estimated as follows:

$$n = \left(\frac{(Z_{1-\alpha} + Z_{1-\beta})x\sqrt{SD_N^2 + SD_O^2}}{d-\mu_D}\right)^2 + 1$$
(24)

Table 4 presents the power $(1-\beta)$ for sample size combination with d = 1, k = 2, α = 0.05, and SD_N=SD_O = 0.4 (for μ of 100 unit). If the required power is 0.80, then the number of samples for new and old procedures should be 18. The 90% CI on the difference of means (\overline{D}) for a paired design used to test equivalence of means was calculated as follows.

$$CI_{\overline{D}} = \overline{D} \pm t_{0.95;n-1} \pm \sqrt{\frac{SD_D^2}{n}}$$
 (25)

$$CI \ of \ SD_{\overline{D}} = \sqrt{\frac{(n-1).SD_{\overline{D}}^{2}}{SD_{O}^{2} \times X_{\alpha:n-1}^{2}}}$$
 (26)

where $X_{\alpha:n-1}^2$ is the percentile from the chi-squared distribution with area α to the left and degrees of freedom n-1.

If SD_0 is not available, then the CI of the SD ratio can be calculated as follows:

$$SD_N = \frac{\sum_{j=1}^n \left(\frac{Y_{jN1} - Y_{jN2}}{\sqrt{2}} - \bar{D}_N\right)^2}{n-1}$$
(27)

$$SD_{O} = \frac{\sum_{j=1}^{n} \left(\frac{Y_{jN1} - Y_{jN2}}{\sqrt{2}} - \bar{D}_{O}\right)^{2}}{n-1}$$
(28)

$$CI of \frac{SD_N}{SD_O} = \frac{SD_N}{SD_O} \sqrt{\frac{1}{F_{\alpha,n-1,n-1}}}$$
(29)

Summary of the comparison method. Accuracy evaluation by using the comparison method has been mentioned by the USP44-NF39 chapter <1225>, BP 2022, and Indonesian pharmacopoeia chapter <1381>[1,2, 4]. The USP 44-NF 39 chapter <1010> [7] described detailed methods for the evaluation of accuracy and precision using the comparison method (Sections internal quality control). BP 2022 [2] used cross-correlation as the method of assessment. However, Indonesian Pharmacopeia [4] did not describe the evaluation method of the accuracy using the comparison method. In comparing two bioanalytical methods, ICH M10 [38] used CCC (Section principle of evaluation). Several applications of CCC in bioanalytical methods have been reported; thus, the application of CCC is recommended instead of using cross-correlations. Further work must be

conducted to determine whether CCC can be applied to replace the complex method described by USP 44-NF39 <1010> [7]. The comparison of two procedures (N and O) should not be evaluated using a significance test with a certain *p*-value; *p*-value cannot be trusted whether it is small or large [41, 42].

Internal Quality Control

Once the proposed method has been validated using separate or combined evaluations as discussed in sections assessment of accuracy and precision-internal quality control, the validated method should always be monitored during its application [9, 43, 44]. A certain number of QC samples should be analyzed during routine analysis. QC samples are typical samples which over a given period are sufficiently stable and homogeneous enough to provide the same results [43]. For each batch of run, 5% QC samples should be analyzed [43, 44]. For every 20 samples, one QC sample should be analyzed. The results of QC sample analysis can be divided into the acceptance zone (between lower and upper limit), guard band, and rejection zone [9]; or limits are set as mean \pm 2 SD, mean \pm 3SD (action limit) [43]; or mean \pm 1 SD (zone C), mean ± 2 SD (zone B), and mean ± 3 SD (zone A) [44]. Based on the general chapter of USP <1210> and <1010>[6, 7], it can be assumed that the acceptance zone is mean \pm CI/PI, whereas the guard band is mean \pm TI. In proving whether the method used is still valid during application, the result of the analysis of all QC samples should be included in the acceptance zone or zones C and B. If the results are in the guard band, action limit, rejection zone, or zone A, then the analytical procedure should be investigated to correct the problem or to be revalidated as necessary. This method can no longer be used for routine applications. The author recommends that the internal quality control methods should be added in the new edition of Indonesian Pharmacopeia [4] and Indonesian's CPOB [45].

Example of Calculations

Determination of %R and recovery curve based on Ref. [1] and [4]. Table 5 shows the analysis result data of an API in a DP (10 level of concentrations) using a UV spectrophotometer; this work aims to investigate systemic and constant proportional errors during extraction. As shown in Table 1, the acceptance criteria for accuracy is 85%–110%, and repeatability is 4% (concentrations of 100 ppm).

 $\overline{\%R} = \frac{\sum_{10}^{1} \%R}{10} = 87.4$, SD = 0.84, RSD = 0.96%; the recovery and precision meet the requirements.

The recovery curve is $X_f = 0.898 X_c - 3.745$, r = 0.999; $Q_{xx} = 18562.5$; $S_y = 1.181$; slope meets the requirement of ICH Q14 [29].

Equation 4: $CI_b = 0.898 \pm t \frac{Sy}{Q_{XX}} (t_{n-2,0.05} = 2.306) = 2.306$. $\frac{1.181}{\sqrt{18562.5}} = 0.898 \pm 2.306 \times 0.00867 = 0.878$ to 0.918, or $0.878 \le b \le 0.918$ (not include 1: a proportional error is observed); Equation 5: $CI_a = 3.746 \pm 2.306 \times 1.181 \sqrt{\left(\frac{1}{10} + \frac{167.5^2}{1856.5}\right)} = 3.746 + 2.306 \times 1.499 = -7.02$ to -0.289; or $-7.02 \le a \le -0.289$, (not include 0: constant systemic error is observed).

These data indicate that the researchers should optimize the extraction methods. The proportional and systemic error of the method cannot be observed using %R, RSD of precision, and slope. The calculation of CI_a , CI_b , and other validation parameters can be performed using our self-developed VMA solutions, which can be downloaded for free using the described link [46].

Determination of Accuracy According to Ref. [6].

Drug substances. Table 6 shows the analysis data of a DS in three levels of API concentration in triplicate using NIR. As shown in Table 2, the acceptance criteria of the bias of the accuracy is -2% to +1%, and the RSD of repeatability and intermediate precision is 1%.

Table 5. Drug product*

Sample	X _c (ppm)	X _f (ppm)	%R
1	100	87	87.0
2	115	100	115.0
3	130	113	86.9
4	145	125	86.2
5	160	141	88.1
6	175	152	86.9
7	190	166	87.4
8	205	181	88.3
9	220	193	87.7
10	235	209	88.5
$\overline{\overline{X}_c}$	167.5		
$\overline{\%R}$ (RSD)			87.4 (0.96)
*Modified from	Dof [28]		

*Modified from Ref. [28]

Table 6.Drug Substance*

Level Concentration	X _c (ppm)	$X_{\rm f} (\rm ppm)$	%R
1	500	498.0	99.6
1	500	494.2	98.8
1	500	498.0	99.6
2	1000	987.2	98.7
2	1000	990.5	99.1
2	1000	999.4	99.9
3	1500	1499.5	99.9
3	1500	1490.5	99.3
3	1500	1481.6	98.8
%R			99.3
RSD			0.48

*Modified from Ref. [6]

Separated evaluation of accuracy and precision. Equation 1: % \overline{R} = 99.32, RSD = 0.49%, Equation 4: CI_b = 0.99380 ± 0.011600; Equation 5: CI_a = -0.58889 ± 12.530 (*p* = 0.05).

Equation 11: $CI_B = (99.3\% - 100\%) \pm 1.86 \text{ x} \frac{0.48}{\sqrt{9}} = -0.7 \pm 0.298 = -0.99\%$ to -0.40%; Equation 12: $CI_{SD} = 0.48 \sqrt{\frac{9-1}{2.73}} = 0.82$. This evaluation indicates that CI_B and CI_{SD} meet the acceptance criteria shown in Table 1 (see 2. 1).

Combined evaluation of accuracy and precision. Equation 13: $\overline{Y} \pm PI = 99.3 \pm 1.86 \times 0.48 \sqrt{1 + \frac{1}{9}} = (99.3 \pm 0.94)\% = (98.4 \text{ to } 100.2)\%$

Equation 14: $\overline{Y} \pm TI (k \times SD) = 99.3 \pm k.0.48 = 99.3 \pm k.0.48 = 99.3 \pm 2.63 \times 0.48 = 98.04\%$ to 100.56% (meet the acceptance criteria, see 2. 2.)

$$k = \sqrt{\frac{Z_{\left(\frac{1+P}{2}\right),n-1}x(n-1)}{X_{\alpha,n-1}^2}} \times (1+\frac{1}{9}), n = 9, P = 0.90, \alpha = 0.10$$
$$k = \sqrt{\frac{1.64^2 x(9-1)}{3.49}} \times (1+\frac{1}{9}) = 2.63$$

k can be obtained using the factors shown in Table XI for TI of the URL of Appendix A Statistical Table [47], whereas $\overline{Y} \pm TI$ can be estimated using the Interactive Statistic Page link [48]. The calculations show that $\overline{\%R} \pm$ PI/TI are included in the specification range (-2% to 1%).

Drug products. Table 7 shows the result of analytical method validation of DP in three runs with different concentration levels, and each run is carried out in six replications. The acceptance criteria of 100 ppm were as follows: accuracy: 85%-110%, bias: -15% to +10%, RSD repeatability: 4% (Table 1).

Accuracy: $\overline{\%R}$ =109.2, SD= 1.40; RSD= 1.27, Recovery curve = X_f = 2.6861 + 1.0646X_c: CIa = 2.6861 ± 4.0840, CIb= 1.0646 ± 0.040306 (p = 0.05, not include 0); CIa = 2.6861 ± 7.7349, CIb = 1.0646 ± 0.7633 (p = 0.01, 0 is included).

This evaluation shows that the recommendation of ICH Q14 [29] regarding the acceptance criteria of slope (0.8–1.25) should be further optimized (P = 0.05 or 0.01; see 2.1.).

Using ANOVA (Equations 7–9), $S_r^2 = 1.6277$; $S_B^2 = 0.42048$; $S_R^2 = 2.0481$.

Table 7. Drug Products*

No of Run (Replication)	X _c (ppm)	$X_{f}\left(ppm ight)$	%R
1 (1)	80	89.9	112.4
1 (2)	80	88.8	111.0
1 (3)	80	87.2	109.0
1 (4)	80	88.4	110.5
1 (5)	80	86.4	108.0
1 (6)	80	88.1	110.1
2(1)	100	109.1	109.1
2 (2)	100	109.2	109.2
2 (3)	100	108.0	108.0
2 (4)	100	108.2	108.2
2 (5)	100	108.0	108.0
2 (6)	100	109.0	109.0
3 (1)	120	132.1	110.1
3 (2)	120	131.6	109.7
3 (3)	120	129.6	108.0
3 (4)	120	128.4	107.0
3 (5)	120	129.6	108.0
3 (6)	120	133.0	110.8
*Unpublished data			

Equation 10: CV mean = $\frac{0.42048}{3} + \frac{1.6277}{3 \times 6} = 0.140 + 0.09$ = 0.23 CI_B = $(109.2 - 100\%) \pm (1.74 \times 1.40/\sqrt{18}) = 10.48 \pm 0.57$

 $CI_{B} = (109.2 - 100\%) \pm (1.74 \times 1.40/\sqrt{18}) = 10.48 \pm 0.57$ = 9.91 to 11.05 % (\alpha = 0.05)

$$CI_{SD} = 1.40 \sqrt{\frac{18-1}{8.67}} = 2.75$$

 $\overline{Y} \pm PI = 109.2 \pm 1.74 \times 1.40 \sqrt{1 + \frac{1}{18}} = 109.2$
 ± 2.50 ; 106.7% to 111.7%

 $\overline{Y} \pm TI \ (k \ x \ SD) = 109.2 \pm 2.19 \times 1.40 = 109.2 \pm 3.07:$ 106.1% to 112.3%

Evaluation of accuracy and precision using the comparison method [7]. The result of the new and validated procedures for homogenous test materials (Section internal quality control) was analyzed using the data presented in Table 8: $n_N = n_0 = 15$ (power = 0.808); $\alpha = 0.05$

$$df = \frac{\left(\frac{0.214}{15} + \frac{0.159}{15}\right)^2}{\frac{0.214^2}{15^2(15-1)} + \frac{0.159^2}{15^2(15-1)}} = 27.4 = 27 \text{ (Equation 22)}$$

$$CI_{\mu_D} = 100.8 - 99.85 \pm 1.703 \sqrt{\frac{0.214}{15} + \frac{0.159}{15}} =$$

[-0.04; 0.50] (Equation 22)

 Table 8. Results of Sample Analysis (n = 15)*

<u>¥</u> (%)	SD^2
$\bar{Y}_{N} = 100.08$	$SD_N^2 = 0.214$
$\overline{Y}_{O} = 99.85$	$SD_0^2 = 0.159$
	$\overline{Y}_N = 100.08$

*Modified from Ref. [7]

$$CI_{SD} = \frac{\sqrt{0.214}}{\sqrt{0.159}} \sqrt{\frac{1}{0.402}} = 1.83$$
(Equation 23)

Non-homogenous test materials with $n_N = n_0 = 18$ (power = 0.803); $\overline{D} = 0.39$; $SD_D^2 = 0.350$; $\alpha = 0.05$; $SD_O^2 = 0.16$ were measured as follows (data from Ref. [7]):

$$CI_{\overline{D}} = 0.39 \pm 1.74 \sqrt{\frac{0.350}{18}} = [0.15 \text{ to } 0.63] \quad (\text{Equation 25})$$

*CI of SD*_{*D̄*} =
$$\sqrt{\frac{(18-1).0.350}{0.16 \times 8.67}}$$
 = 1.81 (Equation 26)

Conclusions and Recommendations

Conclusions and Recommendations: (1) The abovementioned examples show that evaluation using the limits of accuracy and precision (Table 1) described by current USP and FI cannot be used to draw valid conclusions. Although \overline{R} and precision meet the requirements, CIa and CIb could not meet the requirements (5.1 and 5.2.2). The author recommends revising the method of assessment in chapters <1225> of the USP-NF [1] and <1381> of the FI VI [4], that is, "Assessment of accuracy can be accomplished in a variety of ways, including evaluating the recovery of the analyte (percent recovery) across the range of the assay, or evaluating the linearity of the relationship between estimated and actual concentrations". The term "or" should be replaced with "and". (2) As discussed in section selection of SD and the methods of evaluations, the application of a combined evaluation using Equation 14 is recommended in evaluating the accuracy and precision of a proposed/new method. (3) The recovery data must be reported as mean \pm CI/PI/TI instead of mean ± SD/RSD (Sections combined evaluation of accuracy and precision and selection of SD and the methods of evaluations). (4) Further works are needed to determine whether the comparison method using concordance correlations can replace the methods proposed in the USP chapter <1010> [7]. In evaluating the accuracy and precision using the comparison method, a significance test with a certain *p*-value should not be applied (Section summary of the comparison method). (5) In obtaining reliable data at a QC laboratory, the application of internal quality control (Section internal quality control) is strongly recommended. The internal quality control must be included in the new edition of the Indonesian Pharmacopeia.

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References

- USP44-NF39 2021, General Chapters, General Test and Assay, <1225> Validation of Compendial procedure (accessed February 22, 2022), p 1–12, https://online.uspnf.com/uspnf/document/1_GUID-E2C6F9E8-EA71-4B72-A7BA-76ABD5E72964_ 4_en-US
- [2] British Pharmacopeia 2022, Volume 5. The Department of Health and Social Care, The Stationery Office on behalf of the Medicines and Healthcare products Regulatory Agency (MHRA) London, 2021
- [3] European Medicines Agency ICH Q2 (R1) 2006, Validation of analytical procedures: text and methodology, (accessed February 22, 2022, https://www.ema.europa.eu/en/documents/scientific -guideline/ich-q-2-r1-validation-analyticalprocedures-text-methodology-step-5_en.pdf
- [4] Kementriaan Kesehatan RI. 2020. Farmakope Indonesia Edisi VI. Kementriaan Kesehatan RI, Direktorat Jendral Kefarmasian dan Alat Kesehatan. Jakarta.
- [5] The International Organization for Standardization. 2019. ISO 5725-2:2019. Geneva.
- [6] USP44-NF39 2021, General Chapters, General Test and Assay, <1210> Statistical Tools for Procedure Validation (accessed January 23, 2022), p 1–29. https://online.uspnf.com/uspnf/document/1_GUID-13ED4BEB-4086-43B5-A7D7-994A02AF25C8_ 6_en-US
- USP44-NF39 2021, General Chapters, General Test and Assay, <1010> Analytical Data - Interpretation and Treatment (accessed January 22, 2022), p 1–29. https://online.uspnf.com/uspnf/document/1_GUID-5C0818CD-E76F-44B8-B504-C202CA762F2A_5_ en-US
- [8] USP44-NF39 2021, General Notices and Requirement (accessed January 31, 2022), p 1–15, https://online.uspnf.com/uspnf/document/1_GUID-6E790F63-0496-4C20-AF21-E7C283E3343E_8_en-US
- [9] USP45-NF40 2022, General Chapters, General Test and Assay <1220> Analytical Procedure Lifecycle (accessed February 25, 2022, p 1-12, https://online.uspnf.com/uspnf/document/1_GUID-

35D7E47E-65E5-49B7-B4CC-4D96FA230821_2_ en-US

- [10] Yuwono, M., Indrayanto, G. 2005. In Brittain, G. (eds.), Profiles of Drug Substances, Excipients, and Related Methodology, 32th ed. Elsevier Inc. London. pp. 243–258.
- [11] Indrayanto, G. 2012. Validation of Analytical Methods-Updated 2011. In Brittain, H.G. (eds.), Profiles of Drug Substances, Excipients, and Related Methodology, 37rd ed. Elsevier Inc. London. pp. 439–463.
- [12] Indrayanto, G. 2018. Validation of Chromatographic Method of Analysis: Application for Drugs derived from Herbs. In Brittain, H.G. (eds.), Profiles of Drug Substances, Excipients, and Related Methodology, 43rd ed. Elsevier Inc. London. pp. 359–392.
- [13] Indrayanto, G. 2018. Recent Development of Quality Control Methods for Herbal Derived Drug Preparations. Nat. Prod. Commun. 13(12): 1599– 1606, https://doi.org/10.1177/1934578X1801301208.
- [14] Wulandari, L., Idroes, R., Noviandy, T.R., Indrayanto, G. 2022. Application of chemometrics using direct spectroscopic methods as a QC tool in pharmaceutical industry and their validation. In Almajed A.A. (eds.), Profile of Drug Substances, Excipients and Related Methodology, 47rd ed. Elsevier Inc. London.
- [15] Indrayanto, G. 2022. The importance of method validation in herbal drug research, J. Pharm. Biomed. Anal. 214: 114735, https://doi.org/10.101 6/j.jpba.2022.114735.
- [16] Food Drug Administrations Office of Regulatory Affair, ORA-LAB-Manual Volume II. 2020. ORA-LAB.5.4.5. Methods, Method Verification and Validation. https://www.fda.gov/media/73920/down load.
- [17] Association of Official Analytical Chemist. 2019. Appendix K: Guidelines for Dietary Supplements and Botanicals. pp. 1-31, http://www.eoma.aoac.or g/app_k.pdf.
- [18] USP44-NF39 2021, General Chapters, General Test and Assay, <854> Mid-Infrared Spectroscopy (accessed January 25, 2022), p 1–16. https://online.uspnf.com/uspnf/document/1_GUID-21493947-8F57-4FC7-9338FD47C1354A3A_4_en-US
- [19] USP44-NF39 2021, General Chapters, General Test and Assay, <856> Near-Infrared Spectroscopy (accessed January 25, 2022), p 1–16. https://online.uspnf.com/uspnf/document/1_GUID-EC7431B5-E08C-465F-A71D-BD04F30B017F_3_ en-US.
- [20] USP44-NF39 2021, General Chapters, General Test and Assay, <857> Ultra Violet-Visible Spectroscopy (accessed January 26, 2022), p 1–9. https://online.uspnf.com/uspnf/document/1_GUID-4C5C1937-524A-4BED-95E7-384EDE3745E0_3_ en-US

- [21] USP44-NF39 2021, General Chapters, General Test and Assay, <852> Atomic Absorption Spectroscopy (accessed November 25, 2021), p 1–4. https://online. uspnf.com/uspnf/document/1_GUID-7A2010E3-BE15-4F5F-9D2B-FA3C47F69220_2_ en-US
- [22] USP44-NF39 2021, General Chapters, General Test and Assay, <730> Plasma Spectrochemistry (accessed November 25, 2021), p 1–4, https://online. uspnf.com/uspnf/document/1_GUID-0547F508-697A-44F4-B2B0-C9C79A294503_2_ en-US
- [23] USP44-NF39 2021, General Chapters, General Test and Assay, <853> Fluorescence Spectroscopy (accessed February 22, 2022), p 1–6, https://online. uspnf.com/uspnf/document/1_GUID-1996D278-BEC4-40A0-AC53-C8E3B31008D8_1_ en-US
- [24] USP44-NF39 2021, General Chapters, General Test and Assay, <858> Raman Spectroscopy (accessed February 22, 2022), p 1–5, https://online.uspnf.com/ uspnf/document/1_GUID-CF617AE5-647A-4B8A-A5B2-80E06EE7D84B_ 2_en-US
- [25] USP44-NF39 2021, General Chapters, General Test and Assay, <761> Nuclear Magnetic Resonance Spectroscopy (accessed February 22, 2022), p 1–10, https://online.uspnf.com/uspnf/document/1_GUID-B980A615-B451-42FF-AC40-4CB8B64163E3_3_ en-US
- [26] USP44-NF39 2021, General Chapters, General Test and Assay, <736> Mass Spectrometry (accessed February 22, 2022), p 1–6, https://online.uspnf.com/ uspnf/document/1_GUID-1121F1D0-AFE7-4506-94CF-9112714BE4EB_2_ en-US
- [27] USP45-NF40 2022, General Chapters, General Test and Assay, <621> Chromatography (accessed January 11, 2022), p 1–19, https://online.uspnf.com/ uspnf/document/1_GUID-6C3DF8B8-D12E-4253-A0E7-6855670CDB7B_ 2_en-US
- [28] Funk, W., Dammann, V., Donnevert, G. 2007. Quality Assurance in the Analytical Chemistry, 2nd ed. Wiley–VCH Verlag GmbH & Co. KGaA. Weinheim Germany. pp. 12–43; 236–238.
- [29] European Medicines Agency ICH Q 14 2022, Analytical Procedure Development, (accessed September 22, 2022, https://www.ema.europa.eu/ documents/scientific-guideline/ich-guideline-q14analytical-procedure-development-step-2b en.pdf
- [30] Ermer, J. 2015. Methods Performance Characteristics. In Ermer, J., Nethercote, P. (eds.), Method Validation in Pharmaceutical Analysis, 2nd ed. Wiley-VCH Verlag. Weiheim. pp. 73–182.
- [31] USP41-NF36, 2018 General Chapter Test and Assay <1010> Analytical Data and Treatment, United Sates Pharmacopeial Convention, Rockville, MD, pp. 6706-6721
- [32] The Japanese Pharmacopoeia, 17th ed. English version 2016 The Ministry of Health, Labour and Welfare. Japan.
- [33] European Medicines Agency ICH Q2 (R2) 2022, Validation of the Analytical Procedure (accessed

September 22, 2022, https://www.ema.europa.eu/ documents/scientific-guideline/ich-guideline-q2r2validation-analytical-procedures-step-2b_en.pdf

- [34] Ermer, J. 2018. Analytical Target Profile: establishment of precision requirement for assay. J. Pharm. Biomed. Anal. 160: 73–79, https://doi.org/1 0.1016/j.jpba.2018.07.035.
- [35] Furwanti, C., Hendrajaya K., Indrayanto, G. 2020. Simultaneous HPLC Determination of Lidocaine Hydrochloride and Hexachlorophene in a Suppository Product. Media Pharm. Ind. 3(1): 26– 36, https://doi.org/10.24123/mpi.v3i1.2455.
- [36] USP44-NF39 2021, General Chapters, General Test and Assay, <1033> Biological Assay validation (accessed November 17, 2021), p 1–17, https://online.uspnf.com/uspnf/document/1_GUID-952E8C3B-738B-40F8-A552-E14026AC78A9_1_ en-US
- [37] Statistic How to Cross-correlation in Statistic. https://www.statisticshowto.com/cross-correlation/
- [38] European Medicines Agency ICH guideline M10 2019 on bioanalytical method validation, European Medicines Agency (accessed December 12, 2021). https://www.ema.europa.eu/en/documents/scientific -guideline/draft-ich-guideline-m10-bioanalyticalmethod-validation-step-2b_en.pdf
- [39] Real Statistics Using Excel. Lin's Concordance Correlation Coefficient. https://www.real-statistic s.com/reliability/interrater-reliability/lins-concorda nce-correlation-coefficient/.
- [40] Statistic How To. Concordance Correlation Coefficient: Definition & Interpretations. https://www.statisticshowto.com/concordance-corre lation-coefficient/.

- [41] Gandevia, S. 2021. Publications, replication and statistics in physiology plus two neglected curves. J. Physiol. 599(6): 1719–1721, http://dx.doi.org/10.11 13/JP281360.
- [42] Gandevia, S., Cumming, G., Amrhein, F., Butler, A. 2021. Replication: Do not trust your p-value, be it small or large. J. Physiol. 599(11): 2989–2990, http://dx.doi.org/10.1113/JP281614.
- [43] Magnusson, B., Örnemark, U. 2014. Eurachem Guide: The Fitness for Purpose of Analytical Methods–A Laboratory Guide to Method Validation and Related Topics, 2nd ed. Gembloux Belgique. pp. 1–62.
- [44] Food Drug Administrations Office of Regulatory Affair, ORA-LAB-Manual Volume II. 2019. ORA-LAB.5.9 Ensuring the Quality of Test Results. https://www.fda.gov/media/73979/download.
- [45] Badan Pengawas Obat dan Makanan Republik Indonesia. 2018. Peraturan Badan Pengawas Obat dan Makanan Nomor 34 Tahun 2018 Tentang Pedoman Cara Pembuatan Obat yang Baik. Jakarta. pp. 1–431.
- [46] VMA-Solutions. 2021. Version 1.5.1. https://github. com/blue-wind-25/VMA-Solutions/releases/tag/1.5.1.
- [47] Appendix A Statistical Tables And Charts. Table XI Factors for Tolerance Interval. http://www.besseg ato.com.br/UFJF/resources/tolerance_table.pdf.
- [48] Interactive Statistic Page. 2005. Tolerance Intervals for Normal Distribution. https://statpages.info/tolin tvl.html.